

NEW IMAGING AGENTS, PRECURSORS THEREOF AND METHODS OF MANUFACTURE

Background of the Invention

[0001] Short half-lived radioactive compounds are useful as imaging agents to diagnose cancers and abscesses. In the routine production of such imaging agents, safety and production considerations must be taken into account. It is advantageous to minimize contact time while ensuring quick and consistent production of the imaging agents.

[0002] There are generally two types of imaging procedures conducted. In positron emission tomography (PET), two beta rays emitted from the decaying radionuclide are detected. In single photon emission computed tomography (SPECT), one beta ray emitted from the decaying radionuclide is detected. PET provides a more exact location of tumors. However, SPECT is simpler and easier to use, and is therefore used more often.

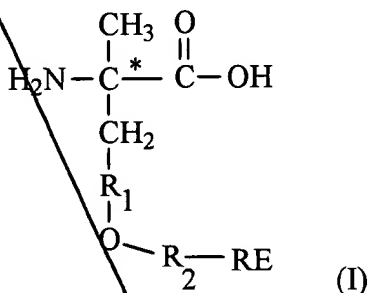
[0003] Generally speaking, PET uses radionuclides that have fewer neutrons than protons, such as ^{18}F , ^{11}C , ^{13}N and ^{15}O , although ^{75}Br and ^{124}I can also be used. SPECT, on the other hand, generally uses radionuclides that have more neutrons than protons, such as ^{67}Ga , ^{77}Br , ^{123}I , ^{124}I , ^{125}I , ^{126}I , ^{131}I and ^{201}Tl .

[0004] In the art, glucose-based and amino acid-based compounds have been used as imaging agents. Amino acid-based compounds are more useful in analyzing tumor cells, due to their faster uptake and incorporation into protein synthesis. Of the amino acid-based compounds, ^{11}C - and ^{18}F -containing compounds have been used with success. ^{11}C -containing radiolabeled amino acids suitable for imaging include, for example, L-[1- ^{11}C]leucine (Keen et al. *J. Cereb. Blood Flow Metab.* 1989 (9):429-45), L-[1- ^{11}C]tyrosine (Wiesel et al. *J. Nucl. Med.* 1991 (32):2041-49), L-[methyl- ^{11}C]methionine (Comar et al. *Eur. J. Nucl. Med.* 1976 (1):11-14) and L-[1- ^{11}C]methionine (Bolster et al. *Appl. Radiat. Isot.* 1986 (37):1069-70). However, ^{11}C -containing radiolabeled amino acids have limited applicability, due to the short half-life of ^{11}C ($T_{1/2}=20$ min.). Therefore, great effort was directed toward the synthesis and evaluation of other radionuclides, such as those listed above. Two ^{18}F -containing radiolabeled amino acids, 4-[^{18}F]fluoro-L-phenylalanine and 2-[^{18}F]fluoro-L-tyrosine, exhibit protein incorporation, but can only be

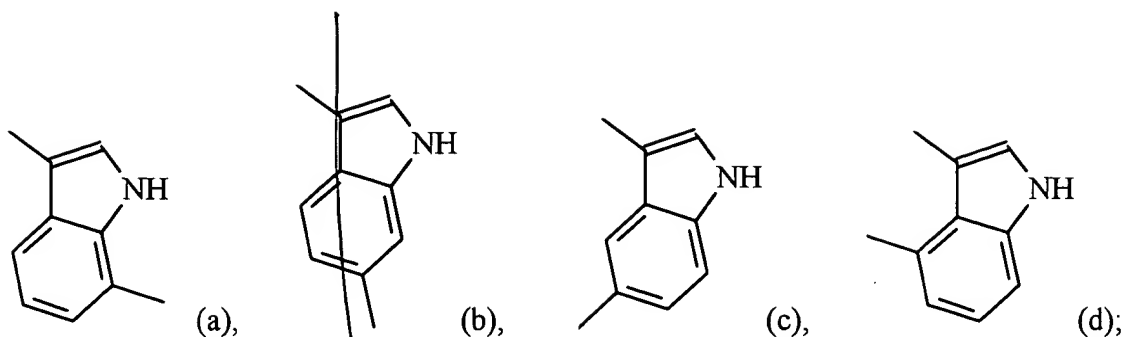
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Summary of the Invention

[0010] The present invention includes compounds of formula (I), or pharmaceutically acceptable salts thereof:



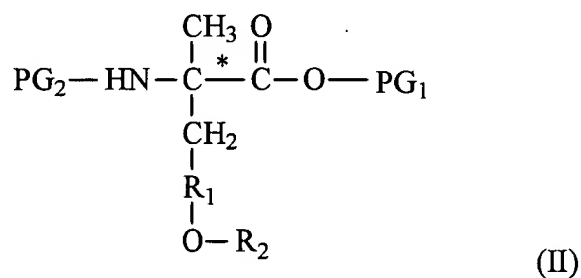
R₁ is selected from the group consisting of a single bond, phenyl, and a group of formula (a), (b), (c) or (d)



R_2 is C_1 - C_7 alkyl; and

RE is selected from the group consisting of ^{11}C , ^{13}N , ^{15}O , ^{18}F , ^{67}Ga , ^{75}Br , ^{77}Br , ^{123}I , ^{124}I , ^{125}I , ^{126}I , ^{131}I and ^{201}Tl , preferably ^{75}Br , ^{124}I or ^{18}F .

[0011] The present invention also includes compounds of formula (II), which are useful for preparing the compounds of formula (I):



wherein

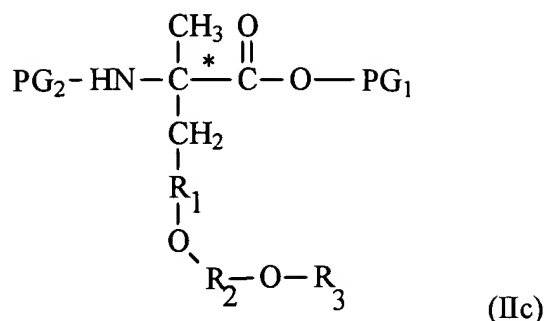
R_1 is the same as indicated for the compounds of formula (I);

R_2 is H or a group $-\text{R}_3-\text{O}-\text{R}_4$, wherein R_3 is C_1 - C_7 alkyl and R_4 is H or a leaving group, preferably a sulfonyl group such as tosyl, trifyl, mesyl, trimsyl, tripsyl, brosyl or nosyl;

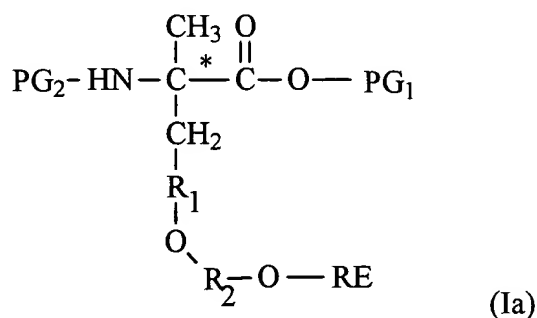
PG_1 is a carboxyl protecting group; and

PG_2 is an amino protecting group.

[0012] The present invention also includes a method of synthesizing compounds of formula (I) by reacting a compound of formula (IIc):



wherein R₁, R₂, PG₁ and PG₂ are the same as indicated for the compounds of formula (I), and R₃ is a leaving group, preferably a sulfonyl group such as tosyl, triflyl, mesyl, trimsyl, tripsyl, brosyl or nosyl, with a salt of RE, where RE is the same as indicated for the compounds of formula I, to produce a compound of formula (Ia):



wherein R₁, R₂, RE, PG₁ and PG₂ are the same as above; and removing the protecting groups.

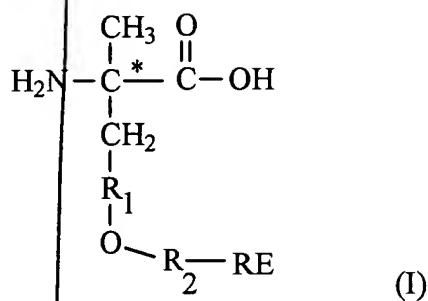
Brief Description of the Drawings

[0013] Figure 1 shows an *in vitro* cellular uptake assay comparison in breast cancer cells between a compound according to the invention (¹⁸F-FPAMT) and a compound of the prior art (¹⁸F-FDG).

[0014] Figure 2 shows an *in vivo* tissue distribution comparison in Fischer 344 tumor-bearing rats between a compound according to the invention (¹⁸F-FPAMT) and a compound of the prior art (¹⁸F-FDG).

Description of the Preferred Embodiments

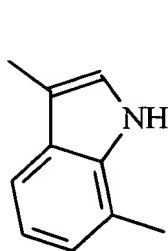
[0015] In this invention, compounds of formula I have been developed:



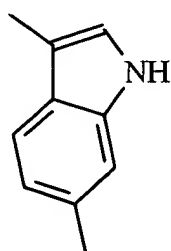
wherein

the C marked with an asterisk represents a chiral center and the compound is present in the L-form, the D-form or as a racemic mixture;

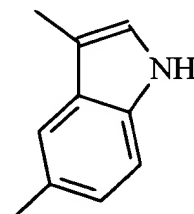
R₁ is selected from the group consisting of a single bond, phenyl, and a group of formula (a), (b), (c) or (d)



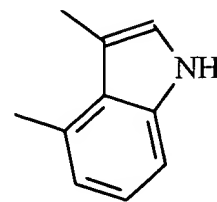
(a),



(b),



(c),



(d);

R₂ is C₁-C₇ alkyl; and

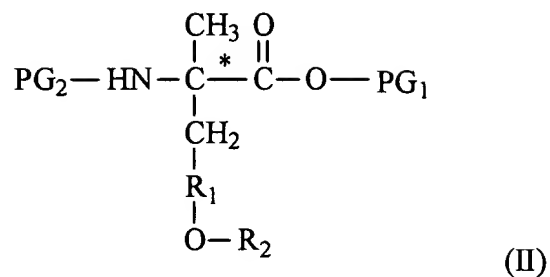
RE is selected from the group consisting of ¹¹C, ¹³N, ¹⁵O, ¹⁸F, ⁶⁷Ga, ⁷⁵Br, ⁷⁷Br, ¹²³I, ¹²⁴I, ¹²⁵I, ¹²⁶I, ¹³¹I and ²⁰¹Tl, preferably ⁷⁵Br, ¹²⁴I or ¹⁸F.

[0016] Preferably, R₁ is phenyl. When R₁ is phenyl, the -O-R₂-RE group can be *para*, *meta* or *ortho* the CH₂ group. It is preferred that the -O-R₂-RE group is *para* the CH₂ group.

[0017] R₂ is preferably C₂-C₆ alkyl, more preferably C₂-C₅ alkyl. It is most preferable that R₂ is propyl.

[0018] The compounds of formula (I) contain a chiral center at the α-C. The compounds of the invention can therefore be present in the L-form, the D-form, or as a racemic mixture. It is preferred that the compounds be present in the L-form.

[0019] The present invention also includes compounds of formula (II):



wherein

R₁ is the same as indicated for the compounds of formula (I);

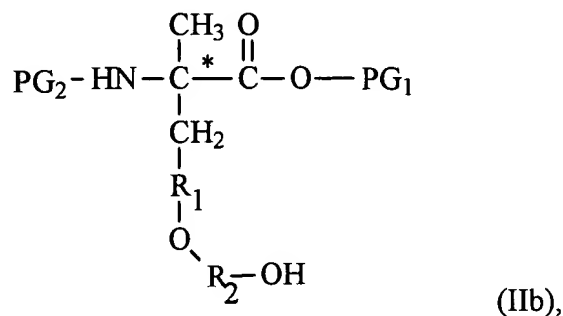
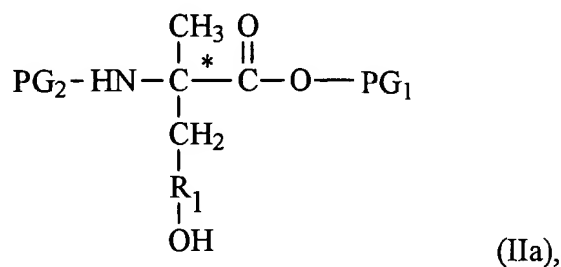
R₂ is H or a group -R₃-O-R₄, wherein R₃ is C₁-C₇ alkyl and R₄ is H or a leaving group, preferably a sulfonyl group such as tosyl, trifyl, mesyl, trimsyl, tripsyl, brosyl or nosyl;

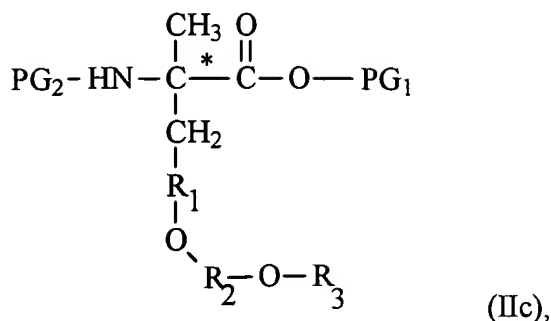
PG₁ is a carboxyl protecting group; and

PG₂ is an amino protecting group.

[0020] The compounds of formula (II) are useful as intermediates in the preparation of compounds of formula (I).

[0021] More specifically, the compounds of formula (II) may be divided into subgroups (IIa), (IIb) and (IIc):





which are important intermediates in the preparation of compounds of formula (I),

wherein in each case

R_1 and R_2 are the same as indicated for the compounds of formula (I);

R_3 is a leaving group, preferably a sulfonyl group such as tosyl, trifyl, mesyl, trimsyl, tripsyl, brosyl or nosyl;

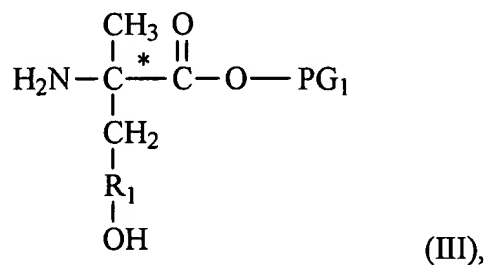
PG_1 is a carboxyl protecting group; and

PG_2 is an amino protecting group.

[0022] As noted above, the leaving groups for compounds of formula (II) are preferably sulfonyl groups, however any suitable leaving group can be used, as known to those of ordinary skill in the art. Especially preferred are tosyl, trifyl, mesyl, trimsyl, tripsyl, brosyl or nosyl, with tosyl, trifyl and mesyl being more preferred and tosyl being most preferred.

[0023] The protecting groups (PG_1 and PG_2) can be any suitable carboxyl protecting group and amino protecting group, as known to those of ordinary skill in the art. Reference is made to "Protective Groups in Organic Chemistry," ed. J. F. W. McOmie, Plenum Press, 1973; and T. W. Greene & P. G. M. Wuts, "Protective Groups in Organic Synthesis," John Wiley & Sons, 1991, each incorporated by reference. PG_1 may be, for example, an alkyl group such as methyl, ethyl or propyl. PG_2 may be, for example, a Boc group.

[0024] Generally, the reaction scheme for preparing compounds of formula (I) is as follows. A starting compound of formula (III):



wherein R₁ and PG₁ are the same as indicated for the compounds of formula (I), such as α-methyl tyrosine C₁-C₃ alkyl ester, α-methyl serine C₁-C₃ alkyl ester or α-methyl hydroxy-tryptophan C₁-C₃ alkyl ester,

is reacted in a suitable solvent with a compound providing a suitable protecting group PG₂, such as di-*t*-butyl dicarbonate (which provides a Boc group), to protect the amine, giving the compound of formula (IIa).

[0025] The compound of formula (IIa) is thereafter reacted in a suitable solvent with a halogenated C₁-C₇ alkanol, to give the compound of formula (IIb).

[0026] The compound of formula (IIb) is thereafter reacted in a suitable solvent with a halogenated leaving group, such as halogen-tosyl, to give the compound of formula (IIc).

[0027] The compound of formula (IIc) is reacted in a suitable solvent with a salt of RE, as defined above, such as K-¹⁸F, to give the compound of formula (Ia). The compound of formula (Ia) is converted into the compound of formula (I) by removing the protecting groups (PG₁ and PG₂), for example by using TFA.

[0028] In the exemplified embodiment, 3-[¹⁸F]fluoropropyl-α-methyl tyrosine, the radiochemical yield for the compound of the invention was much higher than ¹¹C- and ¹⁸F-labeled amino acids synthesized using an electrophilic process. In addition, the compounds of the invention are stable and show high uptake in tumors. The compounds of the invention are useful to differentiate the degree of malignancy of tumors (e.g., brain, breast, prostate, colon, lung, liver, pancreas, gastric, lymphoma, uterine, cervical, extremities, sarcoma and melanoma). The compounds of the invention are also useful to image neurological disorders (e.g. Alzheimer's, Huntington's), abscess, inflammation and infectious diseases. Suitable dosages are known or could be easily determined by those of ordinary skill in the art.

EXAMPLES

Experiment 1. Synthesis of N-*t*-Butyloxycarbonyl-α-methyltyrosine methylester

[0029] α-Methyltyrosine · HCl methylester	MW	245.7
	Wt	2.54 g
	mmol	10.33
Di- <i>t</i> -butyl dicarbonate	MW	218.25

Triethylamine	Wt	2.20 g
	mmol	10.08
	MW	101
	Wt	2.50 g or 3.6 ml
	mmol	24.8

[0030] The α -methyltyrosine methylester (2.54 g) was dissolved in 16 ml of dimethylformamide (DMF) which became cloudy with the addition of triethylamine (3.6 ml). Di-t-butyl dicarbonate (2.20 g) was added and the mixture stirred overnight. After evaporation of DMF under high vacuum the material was treated with ether (30 ml) which produced a precipitate. This precipitate was filtered and discarded. Evaporation of the filtrate yielded 3.38 g (dried under vacuum) of a colorless liquid. TLC analysis using Hexane: Ethyl Acetate (1:1) showed the product at R_f 0.7 and the starting material at 0.1.

Experiment 2. Synthesis of N-t-Butyloxycarbonyl-O-[3-hydroxypropyl]- α -methyltyrosine methylester

[0031] N-t-Butyloxycarbonyl- α -methyltyrosine methyl ester	MW	309.2
	Wt	1.7 g
	mmol	5.50
3-Bromopropanol	MW	138.9
	Wt	924 mg or 0.6 ml
	mmol	6.65
Sodium ethoxide (NaOEt)	MW	68.05
	Wt	742 mg
	mmol	10.90

[0032] The N-t-butyloxycarbonyl- α -methyltyrosine methylester (1.70 g) from the previous synthesis was dissolved in 25 ml of absolute ethanol and treated with dry sodium ethoxide (742 mg). 3-Bromopropanol (924 mg, 0.6 ml) was added and the solution was refluxed for 5 hours. The ethanol was then evaporated, 100ml of dichloromethane added and the solution washed with 100ml of water. After drying over $MgSO_4$ the solvent was evaporated to yield two spots on a TLC plate developed in EtOAc : Hexane 1:1. The first

spot was product and the second was starting material. The product was purified on a silica gel column using EtOAc : Hexane (1:1) A yield of 620 mg (MW 367.2, 31%) was obtained.

Experiment 3. Synthesis of N-t-Butyloxycarbonyl-O-[3-tosylpropyl]- α -methyltyrosine methylester

[0033] N-t-Butyloxycarbonyl-O-[3-hydroxypropyl]- α -methyltyrosine	MW	367.2
Methylester	Wt	620 mg
	mmol	1.69
p-Toluenesulfonyl chloride (tosyl chloride)	MW	190.5
	Wt	400 mg
	mmol	2.10 mg
Pyridine	MW	79
d = 0.98 g/ml	Wt	500 mg
	mmol	6.33

[0034] The N-t-butyloxycarbonyl-O-[3-hydroxypropyl]- α -methyltyrosine methylester (620 mg) from the previous synthesis was dissolved in 10 ml of chloroform and carefully treated with p-toluenesulfonyl chloride (400 mg, which is tosyl chloride) and 0.5 ml of pyridine (500 mg). After allowing to react overnight the volume of solvent was reduced to 2 ml and added to a silica column for purification. The column was washed with EtOAc : Hexane 2:1 and the fractions #4-6 were combine to yield 400 mg, 45.4% of product. Analysis by TLC developed in EtOAc : hexane showed the product between the starting material and tosyl chloride.

Experiment 4. Synthesis of 3- ^{18}F -fluoropropyl- α -methyltyrosine ([^{18}F]FPAMT)

[0035] Fluorine-18 in 1.0 milliliter of 95% oxygen-18 enriched water was transferred from the cyclotron target to an anion exchange column (in the OH form) where the ^{18}F was trapped, and the ^{18}O -water passed through and was collected in a bottle. The ^{18}F was washed from the column with 1.5 milliliters of 0.01 molar potassium carbonate into a reaction vessel containing 26 mg of 2,2,2-Kryptofix. The resulting potassium fluoride/Kryptofix complex was subsequently dried, first by distillation under vacuum

and then by azeotropic distillation of the remaining water with the addition of three 1.5 milliliter portions of acetonitrile.

[0036] The ^{18}F complex was allowed to exchange with the tosyl group on 4 milligrams of N-t-butyloxycarbonyl-O-[3-tosylpropyl]- α -methyltyrosine methylester (tosyl precursor; PAMT-OTs) which was dissolved in 0.5 milliliters of acetonitrile (tube #1). This exchange took 15 minutes at 95°C.

[0037] The cooled acetonitrile solution was passed onto a dry normal 500 milligram silica column (Sep-Pak column (Not C-18)). This material was eluted (washed from the column) with 3 milliliters of diethyl ether into tube #2 where the solvents (ether/acetonitrile) were evaporated to about 1 milliliter.

[0038] Addition of 0.1 milliliter of trifluoroacetic acid for 20 minutes at room temperature was used to remove the protecting group. The solution was evaporated to dryness and 2 milliliters of water were added to this tube (#2). After warming (40°C), the water solution was passed through a 0.22 micron filter for sterilization.

[0039] The final product was analyzed for sterility and by High Performance Liquid Chromatography and TLC for radiochemical purity.

Experiment 5. In vitro cellular uptake assay

[0040] Each well containing 50,000 breast cancer cells, cell line 13762 (0.5 ml/well) was added with 2 μCi of ^{18}F -FDG and ^{18}F -FPAMT. After 0.5-4 hours incubation, the cells were washed with phosphate buffered saline 3 times and followed by trypsin to loosen the cells. The cells were counted by a gamma counter.

[0041] An increased uptake of ^{18}F -FDG and ^{18}F -FPAMT as a function of time was observed in the breast cancer cell line. ^{18}F -FPAMT showed slightly higher uptake than ^{18}F -FDG at 0.5 and 2 hours incubation (Figure 1).

Experiment 6. In vivo tissue distribution

[0042] Fischer 344 tumor-bearing rats (weight 150-155 g) with tumor volume 1.2 cm were administered with ^{18}F -FDG and ^{18}F -FPAMT (10 μCi /rat, i.v.). Biodistribution studies were conducted at 30 min, 2 and 4 hours.

[0043] Tumor/blood and tumor/muscle ratios were increased as a function of time for both ^{18}F -FDG and ^{18}F -FPAMT. There was an increased bone uptake in ^{18}F -FPAMT at 2 hours, suggesting in vivo defluorination might have occurred (Figure 2). Tables 1 and 2 show % of injected dose per gram of tissue weight (values shown represent the standard deviation from 3 animals).

[0044] TABLE 1. Biodistribution of ^{18}F -FPAMT in Breast Tumor-Bearing Rats
% of injected ^{18}F -FPAMT dose per organ or tissue

	30 min	2 h	4 h
Blood	0.638±0.021	0.298±0.032	0.081±0.007
Lung	4.812±0.457	0.541±0.084	0.105±0.008
Liver	6.542±1.142	3.587±0.500	0.930±0.049
Stomach	0.440±0.052	0.190±0.024	0.070±0.002
Spleen	0.404±0.023	0.215±0.029	0.067±0.009
Kidney	8.060±0.101	7.721±1.027	1.467±0.236
Thyroid	0.730±0.126	0.860±0.045	1.220±0.260
Muscle	0.318±0.043	0.154±0.017	0.055±0.012
Intestine	0.838±0.058	0.627±0.209	0.148±0.015
Tumor	0.454±0.016	0.312±0.024	0.069±0.003
Bone	0.447±0.045	1.256±0.117	1.502±0.214
Heart	0.445±0.019	0.219±0.024	0.054±0.007
Tumor/Blood	0.712±0.013	1.079±0.171	0.853±0.036
Tumor/Muscle	1.464±0.138	2.057±0.176	1.322±0.341
Tumor/Lung	0.095±0.006	0.599±0.079	0.658±0.081

[0045] TABLE 2. Biodistribution of ^{18}F FDG in Breast Tumor-Bearing Rats
% of injected ^{18}F FDG dose per organ or tissue

	30 min	2 h	4 h
Blood	0.58±0.044	0.47±0.072	0.25±0.017
Lung	0.62±0.054	0.52±0.137	0.23±0.022

